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Synthesis of the C1–C16 fragment of spirastrellolide A

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Abstract—Synthesis of the C1–C16 fragment of spirastrellolide A is described here featuring Sharpless asymmetric epoxidation, an acid promoted *O*-1,4-addition, and Mukaiyama 1,3-*anti*-aldol. © 2006 Elsevier Ltd. All rights reserved.

We have been working toward a total synthesis of spirastrellolide A, a spiroketal-rich macrolide found recently from marine sponge Spirastrellolide coccinea (Scheme 1).¹ In addition to its ability to cause untimely mitotic arrest in cells, spirastrellolide A was shown to exhibit potent inhibitory activity against protein phosphatase 2A $[IC_{50} = 1 \text{ nM}]$ with an impressive selectivity for PP2A over PP1 [ratio of IC_{50} values = 1:50].^{1,2} Given its biological relevance in cancer therapeutic development, and it structural challenge, spirastrellolide A has already attracted attentions from the synthetic community.³ Retrosynthetically, we planned to approach spirastrellolide A via connecting three major fragments: spiroketal 1, pyran 2, and trioxadispiroketal 3. Recently, we communicated⁴ our synthesis of the C11-epi-C22-C23 fragment (see 1) as a proof-of-concept application





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to feature a ketal-tethered RCM strategy⁵ for constructing of spiroketals.⁶ We report here our synthesis of C1– C16 fragment.

Our synthesis commenced with 1,5-pentanediol, which was quickly transformed into aldehyde 4^7 in 47% overall yield via mono-benzylation and Swern oxidation⁸ (Scheme 2). HEW-Modified Wittig olefination,⁹ followed by Dibal-H reduction, sets up the key allyl alcohol **5** in 95% overall yield exclusively as an *E*-isomer. Sharpless asymmetric epoxidation^{10,11} employing D-(-)-diisopropyl tartrate provided epoxy alcohol **6** in 96% ee,¹² thereby establishing the key C3 stereocenter.

It is noteworthy that we have provided here an alternative approach for establishing the C3 stereochemistry in





spirastrellolide A, as both Paterson^{3a-c} and De Brabander^{3d} employed an asymmetric Brown-allylation. A directed-reductive ring-opening of the epoxide proceeded regioselectively and a subsequent diol protection using 2,2-dimethoxy propane gave acetonide **7**.

With the optically enriched acetonide 7 in hand, we proceeded to complete the pyran synthesis. As shown in Scheme 3, debenzylation followed by a modified-Moffat oxidation¹³ gave aldehyde **8**. Wittig olefination and subsequent hydrolysis of the acetonide group occurred concomitantly with the pyran formation through a thermodynamically controlled intramolecular O-1,4-addition to afford **10** in 91% overall yield, thereby completing the synthesis of C1-10 fragment of spirastrellolide A. The relative *syn* relationship at C3 and C7 was confirmed through NOE. Ensuing protection of the C-1 alcohol in **10** gave the pivaloyl protected methyl ketone **11**, which is suitable for a potential C10–C11 connection as proposed in Scheme 1.

To examine the concept of connecting C10 and C11 through a diastereoselective aldol addition, we prepared aldehyde **12** and **13**.^{14,15} As shown in Scheme 4, by employing Mukaiyama's conditions,¹⁶ methyl ketone **11** was first converted to its respective TMS enol ether using LDA and TMSCl, and the resulting TMS enol ether intermediate was added to aldehyde **12** or **13**, followed by the addition of a stoichiometric amount of BF_3 - Et_2O at -78 °C to give the aldol product **14** in



Scheme 3.



24% yield with a diastereomeric ratio of about 7:1 with the major isomer assigned as the desired 1,3-*anti* product based on Evans' non-chelation 1,3-asymmetric induction model.¹⁷ By using aldehyde **13** under the same conditions, the yield for the respective aldol product **15** was much improved but the ratio was 3:1. On the other hand, by following Evans boron enolate conditions,¹⁸ addition of the di-*n*-butylboron enolate [not shown] derived from methyl ketone **11** with aldehyde **12** in CH₂Cl₂ at -78 °C afforded **14** with 29% yield but in ~1:1 ratio.

We have described here a concise synthesis C1-10 pyranyl unit that differed from the existing syntheses of comparable fragment in spirastrellolide A. We have also demonstrated the feasibility of connecting C10 and C11 through a selective Mukaiyama-type aldol addition, thereby constituting a synthesis of the C1-16 fragment. Efforts are under way in completing the total synthesis of spirastrellolide A.

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- 7. Selected characterizations of new compounds. Compound 5: $R_{\rm f} = 0.35$ [33% EtOAc/hexane]; ¹H NMR (400 MHz, CDCl₃): δ 1.44–1.51 (m, 2H), 1.60–1.67 (m, 2H), 1.80 (s, 1H), 2.04–2.09 (q, J = 7.2 Hz, 2H), 3.47 (t, J = 6.4 Hz, 2H), 4.08 (d, J = 7.2 Hz, 2H), 4.50 (s, 2H), 5.60–5.73 (m, 2H), 7.25–7.37 (m, 5H); ¹³C NMR (125 MHz, CDCl₃): δ 26.0, 29.5, 32.2, 63.8, 70.4, 73.1, 127.8, 127.9, 127.9, 128.6, 128.6, 129.6, 132.9, 138.8; IR (neat) cm⁻¹; 3393br s, 2934s, 2858s, 2361s, 1455s, 1363s. Compound **6**: $R_{\rm f} = 0.10$ [30% EtOAc

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in hexanes]; $[\alpha]_{D}^{23}$ 36.6 [c = 0.89, CH₂Cl₂]; ¹H NMR (500 MHz, CDCl₃): δ 1.54–1.65 (m, 2H), 1.61–1.69 (m, 1H), 1.86 (br s, 1H), 2.91 (dt, J = 4.0, 2.5 Hz, 1H), 3.50 (dt, J = 2.5, 5.5 Hz, 1H), 3.49 (t, J = 6.5 Hz, 2H), 3.62 (ddd, J = 4.5, 7.0, 11.5 Hz, 1H), 3.89 (ddd, J = 2.5, 5.5, 13.0 Hz, 1H), 4.51 (s, 2H), 7.28–7.30 (m, 1H), 7.30–7.35 (m, 4H); ¹³C NMR (125 MHz, CDCl₃): δ 22.7, 29.5, 31.4, 56.0, 58.7, 61.9, 70.1, 72.9, 127.6, 127.7, 128.4, 138.5; IR (neat) cm⁻ 3434br s, 3030m, 2936s, 2861m, 1100s; mass spectrum (APCI): m/e (% relative intensity) 237.2 (M+H)⁺ (100). Compound 7: $R_f = 0.35$ [17% EtOAc/hexane]; ¹H NMR (400 MHz, CDCl₃): δ 1.38 (s, 3H), 1.38–1.43 (m, 2H), 1.44 (s, 3H), 1.46-1.58 (m, 4H), 1.63 (tt, J = 6.8, 6.8 Hz, 2H), 3.47 (t, J = 6.8 Hz, 2H), 3.82 (ddd, J = 1.6, 5.2, 12.0 Hz, 2H), 3.95 (td, J = 2.8, 12.4 Hz, 1H), 4.54 (s, 2H), 7.26–7.37 (m, 5H); ¹³C NMR (125 MHz, CDCl₃): δ 19.5, 21.8, 29.9, 30.3, 31.5, 36.5, 60.3, 69.0, 70.5, 73.1, 98.4, 127.7, 127.9, 127.9, 128.6, 128.6, 138.9; IR (neat) cm⁻¹ 2992s, 2940s, 2862s, 2360s. Compound 8: $R_f = 0.45$ [33% EtOAc/hexane]; ¹H NMR (400 MHz, CDCl₃): δ 1.38 (s, 3H), 1.39–1.44 (m, 1H), 1.45 (s, 3H), 1.47–1.83 (m, 5H), 2.46 (td, *J* = 1.6, 6.4 Hz, 2H), 3.81-3.88 (m, 2H), 3.93-3.99 (td, J = 2.8, 12.0 Hz, 1H), 9.77 (t, J = 1.6 Hz, 1H); ¹³C NMR (125 MHz, CDCl₃): δ 18.0, 19.5, 30.2, 31.5, 36.0, 44.0, 60.2, 68.8, 98.5, 202.8; IR (neat) cm⁻¹ 3300m, 2993s, 2938s, 2869s, 1718s. Compound 10: $R_f = 0.33$ [67% EtOAc/ hexane]; ¹H NMR (400 MHz, CDCl₃): δ 1.17-1.35 (m, 2H), 1.51-1.65 (m, 3H), 1.66-1.76 (m, 1H), 1.81-1.87 (m. 1H), 2.16 (s, 3H), 2.47 (dd, J = 4.8, 16.0 Hz, 1H), 2.57 (br s, 1H), 2.67 (dd, J = 7.6, 16.0 Hz, 1H), 3.60 (dddd, J = 2.0, 3.6, 8.4, 12.8 Hz, 1H), 3.76 (dt, J = 5.6, 5.6 Hz, 2H), 3.83 (dddd, J = 2.0, 4.8, 8.0, 12.8 Hz, 1H);¹³C NMR (125 MHz, CDCl₃): *b* 23.5, 31.1, 31.3, 31.4, 38.2, 50.3, 61.5, 74.2, 78.5, 207.5; IR (neat) cm⁻¹; 3852m, 3675m, 3649m, 3629m, 3376br s, 2934s, 2860s, 2340s, 1716s, 1670s; mass spectrum (APCI): *m/e* (% relative intensity) 187 (M+H, 100), 169 (24), 151(27), 129(84), 111(18). Compound 11: $R_f = 0.40$ [25% EtOAc/hexane]; ¹H NMR (400 MHz, CDCl₃): δ 1.19 (s, 9H), 1.53-1.63 (m, 2H), 1.53-1.63 (m, 3H), 1.75 (dd, J = 7.0, 13.0 Hz, 1.82–1.85 (m, 1H), 2.18 (s, 3H), 2.41 (dd, J = 5.0, 15.0 Hz, 1H), 2.66 (dd, J = 7.5, 15.0 Hz, 1H), 3.42 (dddd, J = 1.5, 7.0, 10.5, 13.0 Hz, 1H), 3.75 (dddd, J = 2.0, 5.0, 8.0, 11.0 Hz, 1H), 4.12 (ddd, J = 6.0, 10.5, 11.0 Hz, 2H); ¹³C NMR (125 MHz, CDCl₃): δ 23.7, 27.4, 27.4, 27.4, 31.2, 31.5, 31.6, 35.6, 39.0, 50.6, 61.3, 74.7, 74.8, 178.7, 207.8; IR (neat) cm⁻¹ 3420br s, 2929s, 2857s, 2360s, 2341s 1710s; mass spectrum (APCI): m/e (% relative intensity) 293 $(M+Na^{+})$ (14), 271 (M+H) (100), 253 (71), 213 (85), 169 (50), 151 (89), 133 (19), 129 (6), 111 (42). Compound 12: $R_{\rm f} = 0.80$ [30% EtOAc in hexanes]; $[\alpha]_{\rm D}^{23}$ 4.35 [c 0.96, CHCl₃]; ¹H NMR (500 MHz, CDCl₃): δ 0.040 (s, 3H), 0.052 (s, 3H), 0.89 (s, 9H), 1.00 (d, J = 7.0 Hz, 3H), 1.62-1.72 (m, 2H), 2.29 (dddd, J = 4.0, 7.0, 7.0, 14.0 Hz, 1H), 3.45–3.51 (m, 2H), 3.77 (ddd, *J* = 4.0, 7.5, 7.5 Hz, 1H), 3.81 (s, 3H), 4.39 (d, J = 11.5 Hz, 1H), 4.44 (d, J = 11.5 Hz, 1H), 4.98 (d, J = 18.0 Hz, 1H), 5.00 (d, J = 10.0 Hz, 1H), 5.77 (ddd, J = 7.5, 11.0, 16.5 Hz, 1H), 6.88 (d, J = 8.5 Hz, 2H), 7.26 (d, J = 8.5 Hz, 2H); ¹³C NMR (125 MHz, CDCl₃): δ -4.51, -4.37, 14.6, 25.9, 33.3, 43.5, 55.3, 67.1, 72.6, 113.6, 114.6, 129.3, 130.7, 140.7, 159.1; IR (neat) cm⁻ 3071w, 2953s, 2856m, 1513m, 1520s. Compound **13**: $R_{\rm f} = 0.65$ (33% EtOAc in hexane); $[\alpha]_{\rm D}^{23}$ +19.3 [c 0.49, CH₂Cl₂]; ¹H NMR (500 MHz, CDCl₃): δ 1.06 (d, J = 7.5 Hz, 3H), 2.45 (ddd, J = 3.5, 4.5, and 16.5 Hz, 1H), 2.50–2.54 (m, 2H), 3.80 (s, 3H), 3.95 (dt, J = 4.5 and 6.5 Hz, 1H), 4.45 (d, J = 11.0 Hz, 1H), 4.53 (d, J = 11.0 Hz, 1H), 5.07 (ddd, J = 1.5, 1.5, and 17.0 Hz, 1H), 5.08 (ddd, J = 1.5, 1.5, and 10.5 Hz, 1H), 5.77 (ddd, J = 7.0, 10.5, and 10.5 Hz) 17.0 Hz, 1H), 6.87 (d, J = 9.0 Hz, 2H), 7.24 (d, J = 9.0 Hz,

2H), 9.75 (dd, J = 2.0 and 3.5 Hz, 1H); ¹³C NMR (125 MHz, CDCl₃): δ 14.2, 40.5, 45.4, 55.5, 71.7, 114.0, 115.9, 129.6, 130.5, 139.9, 159.5, 202.0; IR (neat) cm⁻¹ 2967m, 2836m, 1722s, 1512s, 1060s, 918s. Compound 14: For the major isomer: $R_f = 0.65$ [30% EtOAc in hexanes]; ¹H NMR (500 MHz, CDCl₃): δ 0.082 (s, 3H), 0.107 (s, 3H), 0.90 (s, 9H), 0.99 (d, J = 7.0 Hz, 3H), 1.19 (s, 9H), 1.20-1.23 (m, 2H), 1.44 (dt, J = 3.0, 9.5 Hz, 2H), 1.49–1.61 (m, 4H), 1.74 (dd, J = 2.0, 13.0 Hz, 2H), 1.82–1.85 (m, 1H), 2.37–2.46 (m, 2H), 2.58–2.61 (m, 1H), 2.66 (dd, J = 8.0, 15.0 Hz, 1H), 3.38-3.42 (m, 1H), 3.75-3.80 (m, 1H), 3.94 (ddd, J = 4.0, 4.0, 8.0 Hz, 1H), 4.10 (ddd, J = 6.0, 11.0,11.0 Hz, 1H), 4.13 (ddd, J = 7.0, 10.5, 10.5 Hz, 1H), 4.23 (ddd, J = 4.0, 8.0, 12.5 Hz, 1H), 5.01 (d, J = 9.0 Hz, 1H),5.02 (d, J = 18.0 Hz, 1H), 5.77 (ddd, J = 7.0, 10.0, 17.0 Hz)1H); mass spectrum (APCI): m/e (% relative intensity) $535.5 (M+Na)^+$ (100). Compound 15: For the major isomer: $R_{\rm f} = 0.20$ [25% EtOAc/hexane]; ¹H NMR (500 MHz, CDCl₃): δ 1.03 (d, J = 7.0 Hz, 3H), 1.19 (s, 9H), 1.20-1.32 (m, 2H), 1.48–1.61 (m, 6H), 1.73 (q, J = 6.5, 13.0 Hz, 2H), 1.82–1.84 (m, 1H), 2.38 (dd, J = 5.0, 15.0 Hz, 2H), 2.54-2.57 (m, 1H), 2.57 (m, 2H), 2.64 (dd, J = 8.0, 15.0 Hz)12.5 Hz, 1H), 3.69 (dddd, J = 1.5, 4.0, 4.0, 8.0 Hz, 1H), 3.75 (m, 1H), 4.11 (ddd, J = 7.0, 7.0, 12.5 Hz, 2H), 4.26 (m, 1H),4.51 (d, J = 10.5 Hz, 1H), 4.58 (d, J = 10.5 Hz, 1H), 5.05 (ddd, J = 1.5, 1.5, 7.5 Hz, 1H), 5.06 (ddd, J = 1.5, 1.5, 1.5)17.5 Hz, 1H), 5.81 (ddd, J = 7.5, 10.5, 17.5 Hz, 1H), 6.87 (d, J = 9.0 Hz, 2H), 7.28 (d, J = 8.5 Hz, 2H); ¹³C NMR (125 MHz, CDCl₃): δ 18.1, 27.5, 31.3, 31.3, 31.3, 35.2, 35.4, 39.4, 41.5, 42.8, 44.4, 54.1, 55.2, 59.4, 65.0, 68.8, 76.2, 78.4, 78.7, 83.0, 117.9, 117.9, 118.8, 133.6, 133.6, 135.0, 144.8, 163.3, 182.6, 214.4; IR (neat) cm⁻¹ 3871m, 3751m, 3677m, 3651m, 3494br s, 2956s, 2936s, 2870s, 2342s, 1719s, 1702s, 1616s; mass spectrum (MALDI): m/e (% relative intensity) 541 (M+Na⁺) (100), 518 (M⁺) (4), 409 (4), 321 (10), 273 (39).

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